

CpDNA-based species identification and phylogeography: application to African tropical tree species

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Abstract

Despite the importance of the African tropical rainforests as a hotspot of biodiversity, their history and the processes that have structured their biodiversity are understood poorly. With respect to past demographic processes, new insights can be gained through characterizing the distribution of genetic diversity. However, few studies of this type have been conducted in Central Africa, where the identification of species in the field can be difficult. We examine here the distribution of chloroplast DNA (cpDNA) diversity in Lower Guinea in two tree species that are difficult to distinguish, *Erythrophleum ivorense* and *Erythrophleum suaveolens* (Fabaceae). By using a blind-sampling approach and comparing molecular and morphological markers, we first identified retrospectively all sampled individuals and determined the limits of the distribution of each species. We then performed a phylogeographic study using the same genetic data set. The two species displayed essentially parapatric distributions that were correlated well with the rainfall gradient, which indicated different ecological requirements. In addition, a phylogeographic structure was found for *E. suaveolens* and, for both species, substantially higher levels of diversity and allelic endemism were observed in the south (Gabon) than in the north (Cameroon) of the Lower Guinea region. This finding indicated different histories of population demographics for the two species, which might reflect different responses to Quaternary climate changes. We suggest that a recent period of forest perturbation, which might have been caused by humans, favoured the spread of these two species and that their poor recruitment at present results from natural succession in their forest formations.

Keywords: African rainforest, *Erythrophleum*, Pleistocene forest refuges, species delimitation

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Introduction

Geographic patterns of genetic diversity provide useful information that enables the different factors that have

influenced the history of a species to be determined retrospectively (Petit *et al.* 2003; Heuertz *et al.* 2006; Duminil *et al.* 2009). Whereas many such phylogeographic studies have been conducted on temperate plants (Petit *et al.* 2003), comparatively few studies have been undertaken on tropical species, despite their importance

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for our knowledge of tropical biomes. Owing to their remarkable diversity, it is frequently difficult to identify species in the field when studying tropical plants, although this is a necessary preliminary step before any phylogeographic study can be carried out (Duminil *et al.* 2006; Ley & Hardy 2010). This difficulty is of particular relevance to the study of tree species that have a taxonomically close relationship, the identification of which frequently relies on the characteristics of flowers and/or fruit. Given that these botanical features are absent most of the year and/or difficult to access in the canopy, it is often difficult to distinguish sister tree species on the basis of morphological characteristics (Duminil *et al.* 2006; Koffi *et al.* 2010).

To overcome difficulties in identifying species using only morphological characteristics, a blind-sampling approach—an approach where samples are collected without being identified definitively—followed by assignment of the individuals to species on the basis of molecular genetic analysis has proven to be effective (Duminil *et al.* 2006). However, so far, such approaches have relied on the use of nuclear markers that are highly polymorphic, such as microsatellites, which are often unavailable for the species of interest. An alternative is to rely on DNA markers that use universal primers. In this case, markers in chloroplast DNA (cpDNA) have been shown to be very useful (Hollingsworth *et al.* 2009), but their power to discriminate between species of tropical trees for identification purposes is limited (Gonzalez *et al.* 2009). The limited discrimination might be because interspecific introgression of chloroplasts can occur in species that are closely related taxonomically. As a consequence, in cases where hybridization is a possibility, phylogeographic studies on a given species should include all closely related taxa for which species identification and/or species delimitation is problematic. It is also important to compare information from nuclear and cpDNA because the rate of introgression is typically higher for the latter, which can lead to discordant patterns of species differentiation (Petit & Excoffier 2009). However, in contrast to cpDNA markers, the use of nuclear DNA (nrDNA) was problematic because universal primers were not available. This problem has now been resolved partly by the development of conserved ortholog sets (COSs) of genes (Li *et al.* 2008).

Chloroplast DNA has often been used for phylogeographic studies because sequences can be obtained easily using universal markers and enable historical demographic events to be reconstructed (Petit & Vendramin 2007). Despite the absence of recombination among cpDNA markers, which means that the inferred genealogy represents only a single realization of the coalescent process, their informativeness for phylogeographic

studies has been demonstrated well. cpDNA enables testing the presence of a phylogeographic structure, namely haplotypes from the same population are on average more related than haplotypes from distinct populations, which can indicate long-term population isolation or past fragmentation. For example, with respect to lowland forest-dwelling species, phylogeographic studies can identify possible zones in which species survived (forest refuges) during the last glacial period and routes by which colonization occurred during periods of forest expansion (El Mousadik & Petit 1996; Dutech *et al.* 2000; Chiang *et al.* 2001; Dobes *et al.* 2004; Heuertz *et al.* 2006). Moreover, cpDNA markers can also be used to infer past changes in population demographics, such as population expansion or decline, by means of neutrality tests (Depaulis *et al.* 2005). Therefore, cpDNA markers can be used simultaneously as (i) a tool for species identification (a barcode-like approach; Hollingsworth *et al.* 2009); and (ii) a tool for inferring the population history of related taxa.

The history of vegetation in Central Africa has been analysed mainly from palynological records, retrospective studies of lake levels and studies using techniques that utilize stable isotopes (e.g. Maley 1991; Marret *et al.* 2006; Bonnefille 2007; Cowling *et al.* 2008; Ngomanda *et al.* 2009). In particular, palynological records have revealed that changes in plant communities of varying degrees of abruptness have occurred at different times (Bonnefille 2007). Climatic oscillations between glacial and interglacial periods that occurred during the Pleistocene have played an important role in shaping the structure of ecosystems in Central Africa. The most recent glacial maxima occurred from 160 000 to 130 000 and from 24 000 to 12 000 cal. years BP (Maley 1996). On the basis of the available evidence, arguments have been marshalled to the effect that the climate in the tropics was dry and cool during glacial episodes, which led to an expansion of savannah and/or mountain forest and a fragmentation of lowland tropical rainforests into regions with sufficient moisture to support these forests (mid-altitude mountain regions and river floodplains). In contrast, during interglacial episodes, conditions were close to those of the present, which led to an expansion of forest formations into regions of savannah. Changes in vegetation during the Pleistocene should not be reduced to simple cycles of fragmentation and expansion of forest, given that transitions among different types of forest communities also occurred. However, there is little doubt that climatic oscillations have caused recurrent episodes of fragmentation/expansion among the populations of rainforest species (Flenley 1998). Consequently, we can expect to find signs of the effects of these events on the phylogeography of species.

During the Holocene, around 2500 cal. years BP, the rainforest in Lower Guinea—the region with the highest level of phytogeographical diversity of the African rainforest, which covers an area from Nigeria to the Republic of Congo (White 1979)—was marked by a period of disturbance owing to a southwards shift of the intertropical convergence zone (ITCZ) (Maley 1992; Ngomanda *et al.* 2009). The contraction of the forests that resulted might have triggered the southward migration of Bantu-speaking people who were engaged in agricultural activities, as evidenced by findings that indicate substantial levels of agriculture in Southern Cameroon (van Gemerden *et al.* 2003). This previous anthropogenic disturbance might have produced effects that still influence the species composition of vegetal communities. Such effects might explain the abundance of late secondary species that must have become established in relatively open or disturbed forest, namely species that currently recruit poorly under a closed canopy (Letouzey 1979). Recent demographic changes might also be detectable genetically in the patterns of allelic polymorphism.

In addition to historical factors, environmental heterogeneity can also influence the distribution of plant species and their genetic structure. The distribution of types of forest is correlated strongly with rainfall levels in Central Africa (Letouzey 1979): evergreen forests that are characterized by an abundance of Caesalpiniaceae species are distributed along the coast (Atlantic equatorial coastal forest), whereas semideciduous forests that are characterized by Ulmaceae and Sterculiaceae species are found inland (northwestern Congolian lowland forests).

The study reported herein focused on two sister tree species from the Leguminosae–Caesalpinoideae family, *Erythrophleum ivorense* A. Chev. (syn. *Erythrophleum micranthum* Harms) and *Erythrophleum suaveolens* (Guill. et Perr.) Brenan (syn. *Erythrophleum guineense* G. Don.), that are found in the Lower Guinea region. Given that these two species are difficult to distinguish in the field, they constitute an ideal model to address the problem of identification of species of tropical trees that are highly similar morphologically. Moreover, given that the two species are widespread geographically in tropical Africa and may be ecologically distinct from each other, they constitute an interesting model for the investigation of the influence of historical factors on the rainforest. First, we carried out a comparative analysis of cpDNA and nrDNA polymorphisms and morphological data gathered from *Erythrophleum* samples from the phytogeographical region of Lower Guinea to address the following question: Can the two species be identified retrospectively on the basis of polymorphism of cpDNA? Then, we characterized the spatial distribution

of cpDNA diversity in each species and addressed the following questions. (i) Is there evidence for a phylogeographic structure that might reflect past fragmentation of populations? (ii) Are there geographic areas that show particularly high or low levels of allelic endemism, which might indicate the presence of old populations or a recent colonization event, respectively? Finally, given that these two species of timber tree are known to suffer from poor natural recruitment, we used neutrality tests to examine whether recent changes in population demographics have taken place.

Materials and methods

Description of the species

The paleotropical woody genus *Erythrophleum* is represented by three species in continental Africa [*Erythrophleum ivorense*, *Erythrophleum suaveolens* and *Erythrophleum africanum* (Welw. ex Benth.) Harms], one species in Madagascar (*Erythrophleum couminga* Baill.), one species in Australia [*Erythrophleum chlorostachys* (F.Muell.) Baillon] and one species in China (*Erythrophleum fordii* Oliver). The phylogenetic relationships among these species are not known. The African species are distributed widely on the continent. *E. ivorense* is found in evergreen and semideciduous tropical rainforests; *E. suaveolens* is found in semideciduous tropical forests, forest galleries and dry forests; *E. africanum* grows in woodlands (Aubréville 1970; Vivien & Faure 1985; Hawthorne & Jongkind 2006). As a consequence, these three species should not be found in sympatry, or only rarely. The study reported herein focused exclusively on the two species of timber tree, *E. ivorense* and *E. suaveolens* (the vernacular name commonly used for both species in the area studied is 'tali'). Both species are the source of very poisonous bark that was formerly used widely in Africa in trials by ordeal.

According to Hawthorne & Jongkind (2006), *E. ivorense* has rather shiny leaflets that blacken when dried. The pods generally measure <10 cm in length. Inflorescences are smaller than 1 cm across when the flowers are fully open. The flowers have reddish brown hairs. Each flower has a minute bract, which more or less persists on the inflorescence axis alongside a persistent short pedicel (<1 mm) when the flower has fallen; this leaves an uneven stalk. *E. suaveolens* has leaflets that are not particularly shiny, and they usually turn a green-grey colour when dried. The upper side of the leaflets displays a fine transverse venation. Some pods are more than 10 cm long. Inflorescences are wider than 1 cm with slender pedicels and no bracts. The pedicels are barely persistent on the stalk, and the axis is left almost without scars when the flowers fall.

Erythrophleum species are hermaphroditic. Pollination is insect-mediated in *E. fordii* (Zhu *et al.* 2009), which is probably also the case in the African species. Seed dispersal is probably mainly ballochorous. However, secondary dispersal by primates can occur (Koné *et al.* 2008; Kunz & Linsenmair 2008). Analysis of population structure using the distribution of the diameters of trunks (data not shown) indicates that both species currently experience difficulty recruiting in Cameroon, Gabon and North Congo, which results in a relatively low number of individuals with a trunk of small diameter. Given that both species are light-demanding during their juvenile phase (Kouadio 2009), this lack of recruitment might reflect the natural succession of the forest formations.

Sampling

Plant material (pieces of leaves or cambium) was sampled from 281 saplings or adult trees of *Erythrophleum* in Cameroon, Gabon, the Central African Republic and the Republic of the Congo (Fig. 1 and Data S1, Supporting information) and dried immediately in silica gel to preserve the DNA. The species could not be identified at this stage because fertile material was rarely available. Individuals were grouped together arbitrarily into 59 localities (hereafter called 'populations') on the basis of their geographic coordinates (Data S1, Supporting information). Within these populations, individuals were separated by at least 100 m and at most 10 km. One individual of *E. africanum* that was collected in

Bénin (N-10°209 E-1°206) and one individual of *Calpocalyx dinklagei* that was collected in the Korup National Park (population 2, Fig. 1) were used as outgroups in the phylogenetic analyses.

Fourteen herbarium samples were collected from four of the populations (three from population 41, five from population 26, two from population 11 and four from population 20) and were deposited in the National Botanical Garden of Belgium under the following designations: Ipandi 4, Ipandi 6, Ipandi 15, Doucet G1, Doucet G4, Doucet G11, Doucet G12, Doucet G21, Doucet G42, Doucet G45, Doucet G46, Doucet G47, Doucet G49 and Doucet G50. Additional material was obtained from herbarium vouchers of fertile material from the National Botanic Garden of Belgium (hereafter called 'herbaria from herbarium collections', Data S2, Supporting information), and DNA was extracted successfully from 31 individuals. The sampling date of the fertile herbarium vouchers allowed the flowering period of individuals of *Erythrophleum* to be identified in a given geographic zone. In addition, 793 seeds were gathered from a total of 22 trees from seven populations (Data S1, Supporting information) for morphological analyses.

Morphological traits

When possible, the following morphological characters were assessed for the available herbarium samples: flower length, pedicel length, pilosity of the flowers, presence/absence of persistent pedicels, colour of leaflets when dried and pod length. It was impossible to

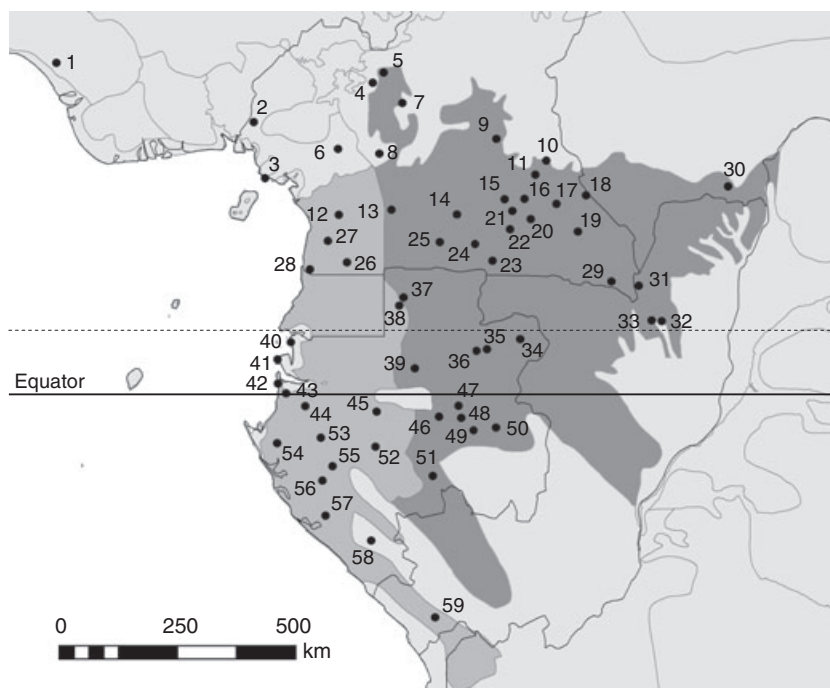


Fig. 1 Map showing locations of study populations (numbers refer to populations as indicated in Data S1, Supporting information). The light-grey zone corresponds to the Atlantic equatorial coastal forest and the dark-grey zone to the lowland forests of northwest Congolian region in accordance with the ecoregions defined by World Wildlife Fund (Olson *et al.* 2001). Details of other ecoregions are not indicated. The position of the equator is indicated as well as the approximate position of the latitude that corresponds to seasonal inversion caused by the annual movement of the intertropical convergence zone (ITCZ, dashed line).

measure flower length and pedicel length when the flowers were immature. We considered only herbarium samples with sufficient discriminatory characters (at least one of the six characters described in Data S2, Supporting information) to enable unambiguous identification of species. Seeds collected in the field were weighed and measured (length of minor and major axes), and the effects of species (classification on the basis of DNA analysis), family, and population were tested using a nested ANOVA with random effects.

Sequencing and genotyping

Total DNA was isolated with a NucleoSpin® plant kit (Macherey-Nagel). Polymorphism of cpDNA was evaluated first by the sequencing of 16 individuals from different populations from Cameroon and Gabon using universal primers. The following six fragments were tested: *psbA/trnH*, *trnC/petN1R*, *trnLR/trnLF*, *rbcl1F/rbcl724R*, *trnT/trnL* and *rpl36/rps8* (Kress *et al.* 2005). Given that the *trnC/petN1R* fragment was the most polymorphic and the other fragments did not provide additional information, only this region was amplified and sequenced using primers described previously in the further analyses (Demesure *et al.* 1995; Lee & Wen 2004). In addition, three chloroplast microsatellites [hereafter called 'ccmps' to distinguish them from the two simple sequence repeat (SSR) motifs present in the *trnC/petN1R* fragment and called '*trnC*-SSRs'] were analysed using universal primers (ccmp2, ccmp4 and ccmp6; Weising & Gardner 1999). Polymerase chain reactions (PCRs) were carried out in a Biometra TProfessional Thermocycler. PCRs (total volume of 25 µL) included 2 µL of template DNA (10–100 ng), 0.1 µL of *Taq* polymerase (Qiagen), 2 µL of PCR buffer, 1 µL of MgCl₂ (25 mM), 0.5 µL of dNTP (10 µM), 0.25 µL of each primer (10 µM) and 18.9 µL of H₂O. The cycling profile for PCR of the *trnC/petN1R* intergenic region included an initial step of 3 min at 94 °C followed by 35 cycles of 30 s at 94 °C, 30 s at 56 °C and 1 min at 72 °C, followed by a final incubation at 72 °C for 5 min. Ccmps were amplified in accordance with the conditions described by Weising & Gardner (1999). Sequences and genotypes were resolved on a 3100 Genetic Analyzer (Applied Biosystems). The lengths of the ccmp fragments were determined by comparison with the GeneScan 350 ROX Size Standard (Applied Biosystems) using the GeneMapper v3.0 software (Applied Biosystems). In general, the *trnC/petN1R* fragment could be amplified successfully for field samples and samples from herbaria that were obtained recently but not from herbaria from herbarium collections. In contrast, the ccmps could be genotyped for all samples, which was probably because

they involve the amplification of relatively short DNA fragments (ca. 90–250 bp). CpDNA haplotypes were defined separately for the *trnC/petN1R* sequences and for size variants at the three ccmps. Subsequent analyses were carried out on a portion or all of the polymorphic characters of *trnC/petN1R* and/or on the ccmp data as detailed below.

The nuclear conserved ortholog set (COS) gene *Eif3E* (Li *et al.* 2008) was amplified to cross-check the results for species identification obtained by the comparison of morphological and cpDNA markers. The PCR protocol was the same as that used for the *trnC/petN1R* sequences except that the annealing temperature was 52 °C. *Eif3E* was sequenced for a subset of the 281 individuals for which the *trnC/petN1R* fragment was amplified successfully. This subset consisted of 93 individuals that represented all cpDNA haplotypes, covered as much of the present study area as possible and included most of the individuals of the populations where both species were found in sympatry. Moreover, one individual of *E. africanum* and one individual of a sister genus, *Erythrophleum sp.*, *C. dinklagei*, were sequenced. Only three haplotypes were obtained for the nuclear *Eif3E* gene for the 93 sequenced individuals of *E. ivorensense* (two haplotypes) and *E. suaveolens* (one haplotype), and only one individual per haplotype was used for the phylogenetic analyses. The accession numbers for the corresponding sequences are as follows: FR670627–FR670631.

Phylogenetic network reconstruction

A minimum-spanning network of the cpDNA haplotypes was reconstructed using MINSPNET software (Laurent Excoffier; available at <http://cmpg.unibe.ch/people/Excoffier-perso.htm>). Given that ccmps may display considerable levels of homoplasy (Provan *et al.* 2001) and that the exact nature of the mutations involved in the size variation of the ccmps was not known, the network was constructed from the *trnC/petN1R* sequences only, using a state difference distance matrix of all polymorphic characters [single-nucleotide polymorphisms (SNPs), insertions–deletions (INDELs) and *trnC*-SSRs]. Note that the *trnC*-SSR microsatellites were used in this analysis, although the ccmp microsatellites were not, because the exact nature of the variation was known for the former (number of repeated motifs). The size polymorphism of ccmps can be because of variation in the number of repeated motifs and/or of the presence of INDELs. The species *E. africanum* was used to root the phylogenetic network.

Maximum-parsimony analysis using a heuristic search with 1000 replicates of random, stepwise branch swapping followed by tree bisection–reconnection was

carried out on the nuclear sequences using the individual of *C. dinklagei* as the outgroup. All characters were set as unordered and having equal weight. One thousand bootstrap replicates using the same search parameters as in the parsimony analysis were produced to assess clade support.

Partitioning of cpDNA variation and testing for a phylogeographic signal

Within-population (H_S) and total (H_T) gene diversities (the probability that two random individuals bear different haplotypes), as well as the level of population differentiation ($G_{ST} = 1 - H_S/H_T$), were estimated separately (i) for SNPs and INDELS from the *trnC/petN1R* sequences; and (ii) for the *ccmp* loci. We also computed the corresponding parameters of diversity measures when distances between haplotypes were taken into account (v_S , v_T , and N_{ST} or R_{ST}) using the method described by Pons & Petit (1996). For the *trnC/petN1R* sequences, the distance between each pair of haplotypes was defined as the number of characters that differed between their sequences (as for the minimum-spanning network), and N_{ST} was estimated using PERMUT software (available at <http://www.pierroton.inra.fr/genetics/labo/Software/Permut/>). For the *ccmp*s, the distance between haplotypes was defined as the squared difference in sequence length, summed over the three *ccmp* loci, and R_{ST} was estimated using CpSSR software (included in the PERMUT software). Populations with fewer than three individuals were discarded for these analyses, following the recommendation of Pons & Petit (1996). We considered that a phylogeographic signal occurred when distinct haplotypes found in the same population were related more closely (i.e. separated by fewer mutational events) on average than distinct haplotypes sampled in different populations, a pattern that results in $N_{ST} > G_{ST}$ and $R_{ST} > G_{ST}$. Such a signal was tested using a randomization procedure of the haplotype assignments in the distance matrices, as implemented in PERMUT software.

On the basis of the results that were obtained earlier in the study, in which individuals were assigned to species by comparing the morphological and genetic data, all genetic analyses were carried out on the whole set of individuals and on two subsets of individuals that putatively matched the two species, *E. suaveolens* and *E. ivorensis* (subsets 'sua' and 'iv').

Geographic patterns of genetic diversity

To evaluate differences in the levels of diversity and genetic structure between the southern (S) and the northern (N) regions of the study area (regions that

were separated by the latitude of seasonal inversion—average latitude of the ITCZ, Fig. 1), as well as between the western (W) and eastern (E) regions (delineated in accordance with the gradient of rainfall), populations were grouped as follows. For *E. suaveolens*, four groups were defined and denoted sua-NW, sua-NE, sua-SW and sua-SE. For *E. ivorensis*, which was found only in the western part of the study area, two groups were defined and denoted iv-N and iv-S. Using SPAGeDi software (Hardy & Vekemans 2002), genetic diversity (H_e) was estimated for all these groups, as well as the ratio of the number of unique haplotypes per group to the total number of haplotypes per group (hereafter defined as N_a). We also measured genetic differentiation (G_{ST} , N_{ST}) and tested for the presence of a phylogeographic signal ($N_{ST} > G_{ST}$) between these groups. These analyses were carried out using a distance matrix (number of polymorphic sites that differed between haplotypes) that was based on the *ccmp* data for *E. ivorensis* (in which no sequence polymorphism was observed, see Results) and on the combination of cpSSRs and *trnC/petN1R* sequences for *E. suaveolens*.

Test of the neutral model

A neutral model with constant population size was tested for each species using the cpSSRs. The frequency distribution of haplotypes is sensitive to demographic changes and selection (Depaulis *et al.* 2005). Gene diversity ($H = 1 - \sum_{i=1}^K p_i^2$, where p_i is the frequency of the i th haplotype) summarizes the frequency distribution of haplotypes, and its expected value can be predicted through simulations for a neutral model with constant population size (null model). An observed value of gene diversity that is lower than expected under the null model may indicate the decline of population size or population structure (or balanced selection), whereas a higher value may indicate population expansion (or a selective sweep, Depaulis *et al.* 2005). To obtain the null distribution of the statistic, we estimated the mutation rate scaled by effective population size ($\theta = 2N\mu$) of each of the five chloroplast microsatellites. The parameter θ was estimated with BATWING (Wilson *et al.* 2003) using a model of constant population size. SNPs and INDELS were included in this analysis only to constrain the trees to compatible topologies in the case of *E. suaveolens*. However, a polymorphic character (SNP4, Data S5, Supporting information) had to be removed because it was not compatible with the hypothesis of no recombination and no recurrent mutation (in SNPs and INDELS) that is used by BATWING. Inferences were obtained from a Markov chain of

2 000 000 steps with a warm-up of 20 000 steps. The prior for θ was chosen to be uniform between 0 and ∞ (results equivalent to a maximum-likelihood framework). Then, 1000 data sets of five linked microsatellites were simulated by a coalescent approach with SimCoal 2 (Laval & Excoffier 2004), using the estimates of θ under a model of constant population size, and the gene diversity H was computed for each data set. The value of H from the empirical data was compared with the distribution of simulated values to estimate a P -value (two-tailed tests). The same analyses were also carried out for the sua-NE and sua-SE subgroups of individuals to check for the existence of a more local demographic signal.

Results

Morphological and phenology observations

Morphological observations of herbarium material. The individuals sampled in coastal forests (Data S2, Supporting information) displayed characters that were typical of *Erythrophleum ivorense*. The flowers had persistent short, c. 0.5-mm-long, pedicels with a minute bract; petals were lanate on their outer side. The mean flower length was 5.3 mm (± 0.8 mm) and mean pod length was 9.9 cm (± 2.8 cm). Leaflets were rather glossy and turned brown upon drying. In contrast, the individuals sampled inland (Data S2, Supporting information) displayed characters that were typical of *Erythrophleum suaveolens* with flowers having a pedicel up to 1 mm long. The petals had sparser and shorter hairs than individuals identified as *E. ivorense* on their outer side and the hairs sometimes covered only the central part or were only present at the margins. The mean flower length was 7.2 mm (± 0.6 mm) and mean pod length was 11 cm (± 1.8 cm). Leaflets were generally glossy and turned green upon drying.

Seed weight. The mean seed weight per family displayed a bimodal distribution (Data S3, Supporting information). The first group comprised eight families from two coastal populations (2, 26), whereas the second group comprised 14 families from five inland populations (9, 16, 18, 20 and 21). In the former, the seed weights ranged from 270 to 450 mg (mean \pm SD = 336 \pm 61 mg), and in the second group, the seed weights ranged from 550 to 800 mg (mean \pm SD = 696 \pm 114 mg). On the assumption that coastal populations belong to *E. ivorense* and inland populations to *E. suaveolens*, nested ANOVA demonstrated that 77% of the variation in seed weight was explained by the species level, whereas the population and family levels explained only 1.3% and 5.7% of the variation, respectively (Data S4, Supporting information).

Flowering periods. Populations in coastal forests flowered from June to October. Populations in inland forests flowered from February to May and from September to November (personal observations and sampling dates of herbarium vouchers of fertile material as described in Data S2, Supporting information).

cpDNA polymorphism

Overall, 19 haplotypes (denoted H1–H19) were defined for the *trnC/petN1R* sequences and 19 haplotypes (denoted A to S) for the three *ccmp* loci (Data S5 and S6, Supporting information). The combination of the two types of marker resolved into 44 haplotypes (Data S6, Supporting information).

The *trnC/petN1R* sequences displayed 17 polymorphic characters among the 281 individuals sampled (12 SNPs, 3 INDELS and 2 microsatellites with length variation, Data S5, Supporting information). Five haplotypes had a frequency of $>2.5\%$ (Fig. 2, Data S5 and S7, Supporting information). According to the phylogenetic network (Fig. 3), H1 was separated clearly from all other haplotypes (by seven mutations). The network was not resolved fully, which indicates the presence of homoplasious characters (SNP4 and *trnC*-SSRs, Data S5, Supporting information). When H1 is omitted from the network, the four most common haplotypes (H2, H4, H10 and H17) are in central positions, which supports the hypothesis that they are more ancestral.

There is a strict association among the haplotypes defined by *trnC/petN1R* sequences and those defined by *ccmps* (Data S8, Supporting information): H1 is associated with K to S whereas H2–H19 are associated with A to J. This association reinforces the phylogenetic distance proposed in the phylogenetic network that clearly separates H1 from all other haplotypes (Fig. 3). The more common haplotypes are also dispersed over a larger area, whereas the rarer haplotypes are located in restricted areas and in general are not shared between populations separated by the latitude of seasonal inversion.

Identification of species using cpDNA and nrDNA

Herbarium vouchers that were identified morphologically as *E. suaveolens* ($n = 26$) always carried haplotype A, E or G at the *ccmp* loci, whereas those identified morphologically as *E. ivorense* ($n = 19$) always carried haplotype K, M, N, O, P, Q or S (Data S2, Supporting information). Haplotypes at the *trnC/petN1R* fragment could only be characterized in the herbarium vouchers that we collected, rather than those collected previously. Samples that were identified morphologically as *E. ivorense* carried haplotype H1, whereas those identified as

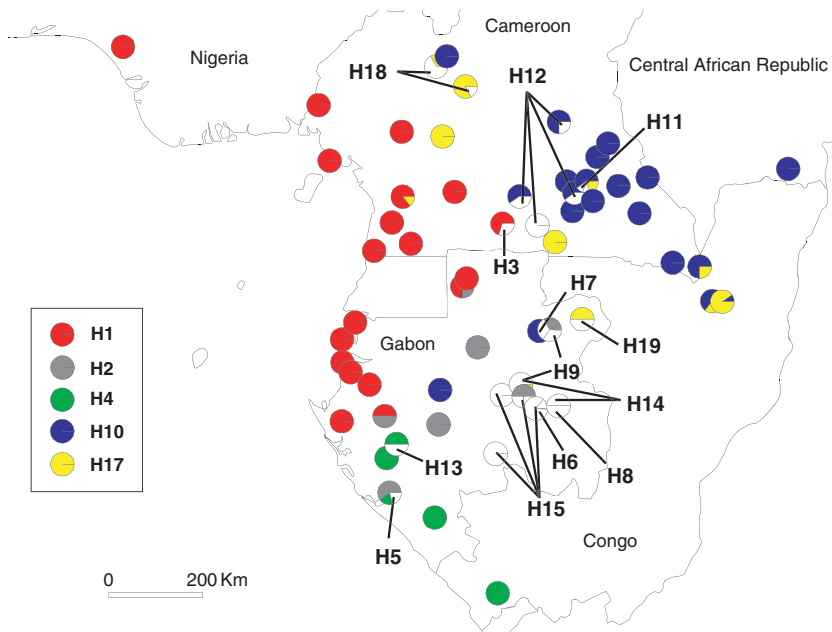


Fig. 2 Relative frequencies of the haplotypes defined by the *trnC-petN1R* intergenic cpDNA fragment in 59 populations of *Erythropheum*. Five haplotypes (H1, H2, H4, H10 and H17) with a frequency $\geq 2\%$ are represented by different colours (see legend). Rare haplotypes are represented as unfilled areas, and their identification number is shown in the map.

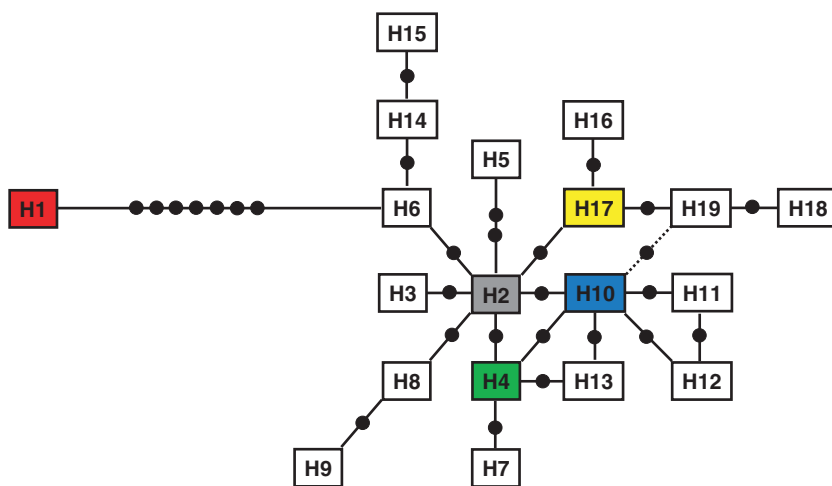


Fig. 3 Minimum-spanning network based on all polymorphic characters within the *trnC-petN1R* intergenic region (SNPs, insertions–deletions, and *trnC*-SSRs). Each black dot on the lines represents one mutation event. The dotted line indicates that H19 was less likely to be derived from H10 than from H17 (H19 and H17 share an insertion, INDEL 1). Information on haplotype frequencies is available in Data S5, Supporting information.

E. suaveolens carried H10. Given that cpDNA is nonrecombinant and there is a perfect association between the haplotypes of the *cmcs* and those of the *trnC/petN1R* fragment (Data S8, Supporting information), our results indicate that individuals with haplotype H1/K–S are *E. ivorensis* and those with haplotype H2–H19/A–J are *E. suaveolens*.

With respect to the nuclear *Eif3E* gene, three haplotypes were defined among individuals of *E. ivorensis* and *E. suaveolens* on the basis of four nucleotide sites that were polymorphic and informative for parsimony analysis (no polymorphic sites that were not informative for parsimony were identified). All individuals with haplotypes H1/K–S formed one monophyletic group and all individuals with haplotypes H2–H19/A–J formed a second monophyletic group (branch support values $>64\%$; Data S10, Supporting information), which

reinforced the identification of species achieved using the cpDNA markers. The phylogenetic reconstruction showed that *E. ivorensis* and *E. suaveolens* were related more closely to each other than they were to *E. africanum*.

Geographical distribution of cpDNA polymorphism

Haplotype H1 (*E. ivorensis*) was restricted to the western part of the geographic area that we studied and, in most coastal populations, was the only haplotype found, except populations in Southern Gabon and Congo. The other four main haplotypes (*E. suaveolens*) were found essentially in the eastern or southern parts of the study area, with two of them present in general in the northern hemisphere (H10 and H17) and the other two in the southern hemisphere (H2 and H4). The

other 14 *trnC/petN1R* haplotypes (*E. suaveolens*) were rare and usually found in restricted geographic areas (Fig. 2). Ten of these rare haplotypes were localized in Gabon and four in Cameroon.

With respect to the three *ccmp* loci, haplotypes A to J (*E. ivorensis*) occurred in the eastern part of the study area, although they were not found in Southern Gabon and Congo, whereas haplotypes K to S (*E. suaveolens*) occurred in the western and southern parts (Data S9, Supporting information).

The two groups of haplotypes (H1/K–S and H2–H19/A–J), and hence the two species, were not usually found in sympatry, with the exception of a few populations that were restricted to transition zones (see Data S1 and S7, Supporting information and Fig. 2).

Genetic structure and diversity analyses for cpDNA

A phylogeographic signal was detected when all individuals from both species were considered ($G_{ST} = 0.69$, $N_{ST} = 0.87 > G_{ST}$, $P < 0.001$ for *trnC/petN1R* sequences; see Table 1 for *ccmps*), namely that haplotypes present within the same population were related more closely on average than haplotypes found in different populations. *E. suaveolens* displayed a phylogeographic signal at the *trnC/petN1R* sequences ($G_{ST} = 0.59$, $N_{ST} = 0.699 > G_{ST}$, $P < 0.01$). In contrast, *E. ivorensis* was monomorphic with respect to the *trnC/petN1R* sequences. At the *ccmp* loci within species, a stronger among-population structure appeared for *E. ivorensis* than for *E. suaveolens* ($G_{ST} = 0.264$ and 0.179 , respectively) and no phylogeographic signal was detected.

For both species, the combination of *trnC/petN1R* sequences and *ccmp* loci revealed a higher genetic diversity in the southern part of the area studied (Gabon) than in the northern part (Cameroon; Fig. 4). For *E. suaveolens*, the highest genetic diversity was found in southeastern populations ($H_e = 0.94$). The genetic differentiation between east and west was $G_{ST} = 0.130$, $N_{ST} = 0.144$ and that between north and south was $G_{ST} = 0.210$, $N_{ST} = 0.130$, with no significant phylogeographic signal detected. For *E. ivorensis*, the genetic differentiation between north and south was $G_{ST} = 0.130$ and $N_{ST} = 0.200$ (test of phylogeographic structure, $N_{ST} > G_{ST}$, $P > 0.05$).

Test of demographic stability

In the test of the neutral model, the observed values of genetic diversity were significantly lower than the simulated values for both taxa (*E. ivorensis*: $\hat{\theta} = 0.5$ (95% CI: 0.20–1.05), $H = 0.766$, P -value = 0.032; *E. suaveolens*: $\hat{\theta} = 1.7$ (95% CI: 1.06–2.70), $H = 0.469$, P -value < 0.0001), which suggested population decline or genetic substructure

Table 1 Genetic diversity within populations (H_s , v_s) and over all populations (H_T , v_T) and population differentiation (G_{ST} , N_{ST} , R_{ST}) estimated for unordered (H , G_{ST}) and ordered (v , N_{ST} , R_{ST}) haplotypes in accordance with Pons & Petit (1996). Numbers in parentheses correspond to standard errors. Analyses were performed at the genus level or within each species, either using the SNPs and INDELs of the *trnC-petN1R* cpDNA intergenic region or using three chloroplast microsatellites (*ccmps*)

	<i>trnC-petN1R</i>											<i>ccmps</i>		
	H_s	H_T	G_{ST}	v_s	v_T	N_{ST}	H_s	H_T	G_{ST}	v_s	v_T	R_{ST}		
<i>Erythrophileum ivorensis</i> /	0.242	0.772	0.687	0.177	1.399	0.873***	0.384	0.658	0.417	0.073	0.756	0.903***		
<i>Erythrophileum suaveolens</i>	(0.0488)	(0.0396)	(0.0540)	(0.0531)	(0.1304)	(0.0383)	(0.0538)	(0.0687)	(0.0485)	(0.0303)	(0.1063)	(0.0381)		
<i>E. ivorensis</i>	†	†	–	–	–	–	0.553	0.752	0.264	0.465	0.730	0.362 ^{NS}		
<i>E. suaveolens</i>	0.294	0.712	0.588	0.169	0.560	0.699**	0.270	0.329	0.179	0.158	0.194	0.187 ^{NS}		
	(0.0618)	(0.0795)	(0.0712)	(0.0463)	(0.0834)	(0.0793)	(0.0605)	(0.0776)	(0.0625)	(0.0419)	(0.0564)	(0.0857)		

*** $P < 0.001$; ** $P < 0.01$; NS, not significant.

†No polymorphism. INDELs, insertions–deletions.

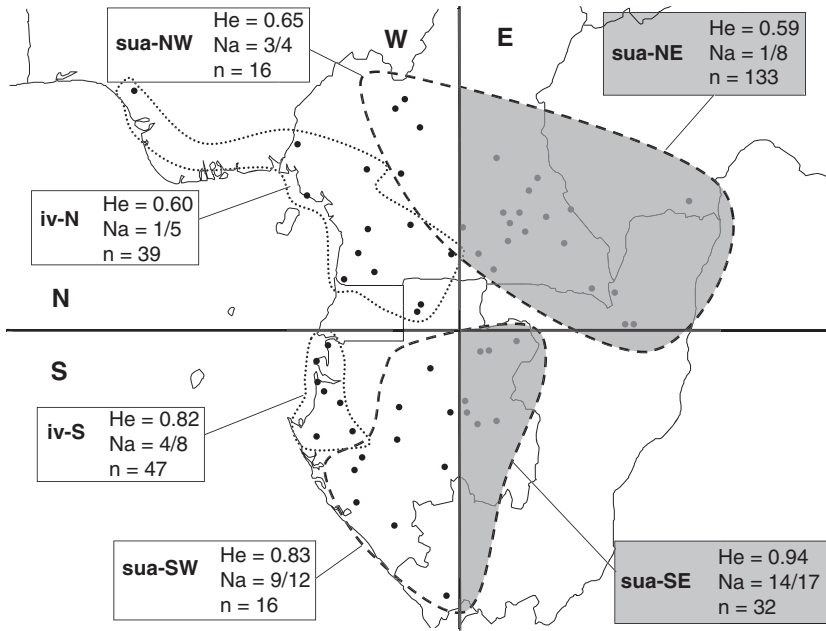


Fig. 4 Diversity (H_e) and number of haplotypes (N_a = number of unique haplotypes/total number of haplotypes) within each of four areas for *Erythrophleum suaveolens* (four zones within the dashed line; two with a grey background, sua-NE and sua-SE, and two with a white background, sua-NW and sua-SW) and two areas for *Erythrophleum ivorense* (dotted lines, iv-N and iv-S). N indicates the number of individuals per zone. The areas are separated by latitude 1.20°N , which corresponds to seasonal inversion, and/or by longitude 12.70°E . The haplotypes are defined by the combination of *trnC/petN1R* sequences and three cpSSR loci.

within each taxon. Analyses on sua-SE and sua-NE were also significant (sua-SE: $\hat{\theta} = 1.0$, 95% CI: 0.42–2.20, $H = 0.75$, P -value < 0.0001 ; sua-NE: $\hat{\theta} = 0.5$, 95% CI: 0.21–1.00, $H = 0.35$, P -value = 0.0004), which suggested population decline.

Discussion

The cpDNA-based approach presented here (Table 2) was effective for the classification of blind-sampled individuals of *Erythrophleum* as *E. ivorense* or *Erythrophleum suaveolens*. Indeed, morphological traits as well as chloroplast and nuclear markers converged to classify individuals near the coast as *E. ivorense* and individuals located inland as *E. suaveolens*. The two species were confirmed to be morphologically distinct (on the basis

Table 2 Available information by type of plant material used in this study

	n^\dagger	Morphology	Short DNA sequence	Long DNA sequence
Herbaria from herbarium collection	31	+	+	–
Herbaria collected during recent field trips	14	+	+	+
Silica gel-dried leaves or cambium	281	–	+	+

† Number of specimens.

of diagnostic characteristics observed in herbarium specimens and weights of seeds gathered in the field) and were shown to display clear genetic differentiation. The use of herbarium vouchers proved to be a very useful approach to complement blind-sampling because it enabled morphological observations to be linked with DNA classification of the individuals. The herbaria that were used in this study were composed of new samples collected in a few localities and herbaria from herbarium collections from a wider geographical area. As summarized in Table 2, the herbaria from herbarium collections were very valuable for the study of morphological characters, but DNA degradation is often a limiting factor for molecular analyses: short cpDNA fragments (100–250 bp for the *ccmp*s studied here) could be amplified, but longer and more informative sequences could not (~1000 bp for *trnC/petN1R* sequences or ~650 bp for *Eif3E* sequences). However, the fact that cpDNA does not undergo recombination should ensure a strong association between haplotypes identified at different loci; indeed, we identified two matching categories of haplotypes using *trnC/petN1R* sequences and *ccmp* loci that were shown to distinguish the two species. Hence, our strategy of using both short and long cpDNA fragments allowed us to determine taxonomic links between herbaria from herbarium collections and blind-sampled individuals (Table 2). This DNA barcode-like approach is very useful in cases where it is difficult to identify individuals in tropical forest taxa. Chloroplast DNA markers appeared to be informative to distinguish *E. ivorense* and *E. suaveolens*. However, one cannot exclude the possibility that

introgression has occurred by hybridization between the two species, particularly in populations where both species are found in sympatry, on the basis of comparisons of morphological characters and cpDNA markers alone. To investigate this issue further, in the absence of available microsatellite markers, the nuclear gene *Eif3E* was sequenced and the identified haplotypes of *Eif3E* associated perfectly with the haplotypes of the cpDNA markers, which supported the hypothesis that the two species were completely isolated reproductively.

However, it must be noted that the plant model used here falls into the category of ideal cases for this type of study (the information provided by morphological and molecular markers agreed strongly in terms of distinguishing the species). A recent review of comparisons between morphological and molecular markers for species delimitation demonstrated that complete congruence was reported in only approximately one-third of the species studied (Duminil & Di Michele 2009). For another one-third of species, morphological markers provided better discrimination than molecular markers, which might indicate labile properties such as phenotypic plasticity, local adaptation or even neutral polymorphism associated with morphological markers. For the remaining species, molecular markers provided better discrimination than morphological markers. This is generally the case when no divergence at all is observed for morphological markers but two or more species can be distinguished with molecular markers (i.e. cryptic species).

Erythrophleum ivorensense and *E. suaveolens* mainly exhibited parapatric geographic distributions, which might reflect their different ecological requirements. In the Lower Guinea region, *E. ivorensense* was generally associated with both higher levels of rainfall (more than 1800 mm per year) and a shorter dry season than *E. suaveolens*, which matches the distribution of the two species in West Africa (Aubréville 1959). Given that the third *Erythrophleum* species that is present in continental Africa, *Erythrophleum africanum*, is found in savannah, all three species could have specialized along a rainfall gradient, which would suggest an ecological mode of speciation. As supported by the analysis of cpDNA (*trnC-petN1R* intergene) and nrDNA (*Eif3E* gene), it is most likely that *E. suaveolens* and *E. ivorensense* are sister species, whereas *E. africanum* is related more distantly. Therefore, the speciation event between *E. ivorensense* and *E. suaveolens* is probably the most recent in continental Africa within the *Erythrophleum* genus.

Erythrophleum suaveolens and *E. ivorensense* presented different distributions of genetic variation. It is difficult to compare the two species with respect to genetic diversity because they occupy parapatric geographic areas: *E. suaveolens* is found mostly in semi-deciduous forests

and *E. ivorensense* mostly in evergreen forests. Both types of forest were probably affected in different ways by historical events, in particular climatic events: evergreen forests probably declined more than semi-deciduous forests during the drier periods of the Pleistocene that were concomitant with the glacial maxima. *E. ivorensense* was monomorphic for the *trnC/petN1R* sequences, whereas substantial polymorphism was found in *E. suaveolens*. Given that there is no obvious difference in life history traits or in generation time between the two species, it is difficult to explain the differences in their levels of diversity on the basis of these factors. The lower diversity in *E. ivorensense* might result from an historically lower effective population size. It might also indicate recent colonization by *E. ivorensense* with founder events. Both these conjectures are compatible with the apparently higher level of rainfall that is required by *E. ivorensense*, which would have suffered a more drastic reduction in population during the cooler and drier periods of the Pleistocene than *E. suaveolens*. Nevertheless, *E. ivorensense* and *E. suaveolens* display comparable levels of cpDNA polymorphism at the *ccmp* loci. If we assume that the rate of mutation is higher at the *ccmp* loci than at the *trnC/petN1R* fragment, an ancient bottleneck in *E. ivorensense* might have caused a persistent loss of diversity at the *trnC/petN1R* locus and a recovery of diversity at the *ccmp* loci. Unfortunately, the *ccmp* loci were typically too homoplasious to infer phylogeographic patterns (uninformative phylogenetic network, data not shown).

The demographic history of *E. suaveolens* can be interpreted in more depth than that of *E. ivorensense*. The spatial distribution of the 18 haplotypes that were defined for the *trnC/petN1R* sequences was very informative. Given their central position in the haplotype network and their relatively high abundance in the Lower Guinea region, the four haplotypes H2, H4, H10 and H17 should have a less recent origin than the others. They have different geographic distributions, H2 and H4 being located mainly in Gabon, and H10 and H17 being located mainly in Cameroon. Moreover, a phylogeographic signal was detected, which points to the effects of previous historical events. We observed a set of rare haplotypes that were endemic to small geographic areas. These rare haplotypes are probably of recent origin and have not yet dispersed into larger areas. The southeastern part of Gabon had a particularly large number of rare haplotypes, which suggests that *E. suaveolens* has remained relatively abundant in or near this area for a period of sufficient length to accumulate new mutations. This region of high diversity is not far from the Chaillu Mountains, which have been proposed to be a forest refuge (Maley 1996). However, the species might also have maintained important

populations in forest galleries. This conjecture is supported by the current distribution of the species in West Africa, where it is typically found as relic populations in forest galleries that are surrounded by savannah. In Central Africa, it is possible that many refuge zones were dispersed throughout the region at low-altitude regions of mountains or along rivers and that the history of fragmentation/recolonization involved frequent admixture. Areas that display a high level of genetic diversity might correspond to areas with long-standing populations or to areas in which different lineages from different refugia have mixed (as suggested by the example of tree species in Europe, Petit *et al.* 2003). In Gabon, the high level of diversity of *E. suaveolens* is constituted mostly of endemic and sometimes closely related haplotypes, which supports the hypothesis of long-standing populations. In contrast to the pattern found in Gabon, southeast Cameroon is dominated by a single haplotype, which suggests that colonization by *E. suaveolens* occurred much more recently in this region. It might reflect a recent expansion of forest cover in southeast Cameroon. Additional data, especially phylogenetically informative data, are needed to explore this issue in more depth.

For both *E. ivorensis* and *E. suaveolens*, the level of genetic diversity was much higher in Gabon than in Cameroon, especially in terms of the number of endemic haplotypes. The north–south divide observed for *E. suaveolens* is also apparent for *Greenwaniodendron suaveolens* (Dauby *et al.* 2010) at cpDNA and for *Distemonanthus benthamianus* (Debout *et al.* 2010) at nuclear markers, which suggests that similar factors might have acted on different species of forest trees. In particular, this divide can be related to physical conditions that have been more favourable historically to tropical forests in Gabon than in Cameroon. First, inland Cameroon and Gabon experience different climates, especially during the main dry season (Nicolas 1977; Suchel 1990; Leroux 1993; Makanga & Boko 2000). In Gabon, the dry season in the south (June–August) is characterized by constant cloud cover and the absence of rainfall, which generate lower temperatures and higher levels of relative humidity than those in Cameroon. Low temperatures, low levels of evapo-transpiration and the existence of morning dew and fog reduce the potentially negative effect of this dry season on vegetation. In contrast, the dry season in Cameroon (December–February) is more stressful for the vegetation, because it is sunnier, with higher temperatures and lower levels of relative humidity. Second, the influence of cooler and drier masses of air from the northern hemisphere is noticeable in Cameroon, but not in Gabon, and is assumed to have had significant effects during the Pleistocene (Leroux 1993). Finally, attempts to recon-

struct changes in vegetation on the basis of climate also provide support for the hypothesis because they indicate that the drier periods of the Pleistocene and Holocene had less impact on the fragmentation of species of forest trees in Gabon than in Cameroon (Anhuf *et al.* 2006). It is likely that all these patterns are related to past climatic events in the Pleistocene era, whereas the current poor recruitment observed in the two species is probably related to more recent events.

The poor recruitment of *E. ivorensis* and *E. suaveolens* might well mirror the recent history of disturbance in the tropical lowland forests of Lower Guinea. These species are post-pioneer light-demanding species that recruit poorly under dense forest cover (Kouadio 2009). The high level of disturbance in the forests some centuries ago might have favoured the spread of *Erythrophloeum* species that have since begun to decline as the forest has become re-established (van Gernerden *et al.* 2003). This conjecture is supported by the presence of charcoal in southeast Cameroon that dates back to between 2000 and 200 BP (region of Mindourou, J.-L. Doucet, data not shown), which could indicate earlier slash-and-burn farming. For both species, signs of a historic decline in population size are apparent, which is somewhat surprising given the relative abundance of individuals with a trunk of large diameter. It is not obvious whether these findings reflect the poor recruitment of the species after past disturbances (climatic and/or human) of the forests during the last few thousand years (van Gernerden *et al.* 2003), because only relatively drastic changes in population size would be detectable over only a few tens of generations, and the population density is still relatively high. Given the risk that our model is flawed because of the effects of natural selection or nonconventional modes of mutation on the molecular markers used, evidence from multiple nuclear markers is needed to confirm whether these populations have been in decline.

Conclusion

We have shown that the approach proposed herein is effective for the classification of blindly sampled individuals into two species. The approach is only feasible if fertile herbaria are available, which highlights the importance of herbarium collections and the need for new collections in tropical zones in which limited prospecting has been undertaken, and where new species might well be found. Our analysis of cpDNA polymorphism in *Erythrophloeum* revealed that different factors shaped the geographic distribution of diversity in these species. First, an ecological (rainfall) gradient seemed to be the main factor that explained the distribution of the cpDNA haplotypes, because the two species, which had

different ecological requirements, bore distinct haplotypes and showed essentially parapatric distributions along an east–west axis. Second, it is likely that historical demographic changes explain the unequal north–south distribution of diversity within each species and the detection of a phylogeographic signal in *Erythrophleum suaveolens*. In particular, the much higher levels of diversity and the prevalence of allelic endemism in the south (essentially Gabon) might be indicative of populations with a less recent origin in this region, which could be related to the presence of Pleistocene forest refuges. The low diversity in southeast Cameroon might indicate relatively recent colonization, which could indicate a concomitant expansion of the forest cover during the Holocene. The signal of population decline, which is difficult to interpret and potentially subject to methodological flaws, must be confirmed using nuclear genes. The role of past human disturbance on forest dynamics requires further analyses, particularly using highly polymorphic nuclear markers.

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The researchers involved in this publication have collaborated for many years in studying the ecology and population genetics of tropical tree species from tropical African rainforests. This collaboration provides an important contribution to the understanding of tropical rainforest history in Africa according to climate changes and anthropogenic influence. Results will allow fundamental application in biodiversity conservation and management.

Supporting information

Additional supporting information may be found in the online version of this article.

Data S1 Characteristics of the 59 populations of *Erythrophleum ivorense* (iv) and *E. suaveolens* (sua) sampled in the Lower-Guinea phytogeographic domain.

Data S2 Morphological and genetic identification of fertile herbarium vouchers.

Data S3 Distribution of mean seed weight per family for 22 trees.

Data S4 Nested ANOVA and variance component estimation for mean seed weight per family.

Data S5 Description and frequencies of the 19 haplotypes identified in *Erythrophleum* by sequencing the chloroplast *trnC/petN1R* intergenic region.

Data S6 Description and frequencies of the 19 haplotypes identified in *Erythrophleum* by genotyping the chloroplast microsatellites *ccmp* 2, 4 and 6.

Data S7 Color figure that integrates Figs 2 and 3 of the manuscript.

Data S8 Number of *Erythrophleum* individuals bearing each possible combination of *trnC-petN1R* and *ccmp* haplotypes.

Data S9 Distribution map of genetic diversity in *Erythrophleum* at *ccmp* loci. Seven haplotypes with a frequency of at least 2% are represented by different colours (legend).

Data S10 Phylogenetic analyses of the nuclear *Eif* fragment.

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